Papers and Originals

Rubella Vaccine Trial in Children

I. B. HILLARY,* M.B., D.P.H., D.C.H.; P. N. MEENAN,† M.D., F.C.PATH.; A. H. GRIFFITH,‡ M.D., D.P.H. C. C. DRAPER,§ D.M., D.P.H., D.T.M.&H.; G. D. LAURENCE,|| B.SC.

British Medical Journal, 1969, 2, 531-532

Summary: A study with RA 27/3 attenuated rubella virus vaccine (Plotkin strain) showed that this produced a significant antibody response in all of twenty-one vaccinated non-immune children without any appreciable marked clinical reactions. Serological examination of 53 non-immune and 29 immune siblings living in the same households failed to show any evidence of transmission of infection.

Introduction

A live attenuated rubella virus strain cannot be regarded as suitable for vaccine manufacture unless, when administered to susceptible subjects, it consistently produces satisfactory titres of rubella antibodies, does not generate undesirable clinical responses, and does not produce infection in susceptible seronegative contacts. Preliminary studies indicate that four strains of attenuated rubella virus may meet these criteria. They are the HPV-77 virus (Meyer et al., 1966; Buynak et al., 1968), the Cendehill strain (Huygelen and Peetermans, 1967; Martin du Pan et al., 1968), the RA27/3 or Plotkin strain (Plotkin et al., 1967, 1968), and the Benoit strain (Buynak et al., 1968). The first three have already been administered to adults in Britain (Dudgeon et al., 1969). This paper reports on clinical experience with the RA27/3 strain in the first rubella vaccine trials in children in Britain and Ireland.

Since at the time of this trial it could not be assumed that rubella-susceptible contacts would not be infected by a vaccine, it was necessary to take precautions against infection of pregnant women during the four to five weeks after vaccination. On the other hand, it was necessary to have rubella-susceptible persons in constant close contact with vaccinees in order to study the contagiousness of vaccine infection. To conform with these requirements the trial was carried out in a rural area of about 300 square miles (777 sq. km.) in Ireland. This area was deliberately chosen because it contained many large families with young children living on relatively isolated farms and with minimal contact between these and other families in the surrounding neighbourhood, especially during school holidays. Also it was felt that the family was the most appropriate unit in which to study the possible spread of virus from a vaccinated child to other siblings or contacts.

- College Graduate, Department of Medical Microbiology, University College, Dublin.
- † Professor of Medical Microbiology, University College, Dublin.
- ‡ Clinical Research Division, Wellcome Research Laboratories, Beckenham, Kent.
- § Department of Virology, Wellcome Research Laboratories, Beckenham, Kent. At present Senior Lecturer, London School of Hygiene and Tropical Medicine, London W.C.1.
- || Research Graduate, Department of Virology, Wellcome Research Laboratories, Beckenham, Kent.

Methods and Materials

Study Population.—The purpose and procedure of the investigation were initially explained in detail to 49 mothers of large isolated families at their homes. Thirty agreed to participate in the study. Blood samples were taken originally only from the mothers in these families, and the sera titrated for haemagglutination inhibition (H.A.I.) antibodies by the filter paper disc method (Draper and Kelly, 1969) and the standard method (Stewart et al., 1967). Three mothers were found to be seronegative and their families were therefore excluded from the trial. Blood samples were then collected from the 150 children between 1 and 17 years of age in the remaining 27 families, but the H.A.I. tests indicated that in some families most or all of the children were immune to rubella. Eventually 19 families with a total of 104 children participated in these studies.

Vaccine.—The RA27/3 strain of rubella virus at the 25th passage level was received from Dr. S. Plotkin and was passaged twice at the Wellcome Research Laboratories to produce vaccine at the 27th passage level. To prepare the vaccine WI-38 cell cultures at the 30th passage level were inoculated with seed virus and incubated at 30° C. for one day. The fluid on the cells was then changed, the cultures were washed twice to remove calf serum, and the cultures were maintained on Eagle's basal medium with serum at 30° C. Multiple harvests made at five and eight days were pooled, stabilized, and freeze-dried in 1-ml. and 0.5-ml. volumes. The cells maintained their normal karyology, were not oncogenic to hamsters, and there were no adventitious agents detected in the vaccine. Ampoules of vaccine freeze-dried from 1-ml. and 0.5-ml. volumes contained 10^{3,2} and 10^{2,9} TCID₅₀ of virus respectively.

Results

Clinical Evaluation—Twenty-one non-immune girls and one immune girl from these 19 isolated rural families were vaccinated with 103 TCID₅₀ of the Plotkin strain rubella virus vaccine. These girls ranged in age from 15 months to 8 years, and in general were the youngest non-immune females in each family. Fifty-three of the siblings in the trial were nonimmune; 29 were immune (Table I). Each vaccinated child had at least one non-immune sibling living in the same house, except in families H and I, where the vaccinee lived in one house and her non-immune contact was a very close friend and playmate in a neighbouring dwelling a few yards away (Table II). All the vaccinated children were examined clinically between Day 8 and Day 12 after receiving the vaccine. Mild enlargement of the postauricular glands was noted in 13, and two had a transient rash between Day 8 and Day 10 (Table III). Symptoms encountered during the 21 days' postvaccination period included arthralgia of the hip in a girl aged 8 years.

She complained of hip pain on Days 8 and 9, and though it was sufficient to make her limp it did not cause any anxiety. The parents clearly attributed the pain to vaccination. Five other children complained of mild malaise on one day only, but otherwise no symptoms were noted among the vaccinees during the 21 days.

TABLE I

No. of Families	No. of Children	Vaccinated	No. of Unvaccinated Siblings			
	Non-immune	Immune	Non-immune	Immune		
19	21	1	53	29		

TABLE II.—Age and Immune State of Vaccinees and Siblings

Fam-	Age in Years															
Fam- ily	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16
ABCDEFGH	- - 0	0	0	_	0 +	<u>-</u> -	0 +	+ - + 0	+ -	_	-		_			
H I	0		0	_		+	+	+	+	+	+	-		+	+	
		0	0	0	-	=	_	0	-		_	+	+	+		
JKLMNOP QR	- 0	0 0	+	_	0+	0 - Ø		+ + +	_	++-	-	+	+	+-+	+	+
S	0 -		-	_			_	-								

^{- =} Non-immune contact. 0 = Immune vaccinee. 0 = Non-immune vaccinee. - = + = Immune contact. T = Twins.

TABLE III.—Clinical Reactions in 21 Non-immune Children Given RA27/3 Rubella Vaccine

Nature	No. of	Mean Day	Mean		
of Reaction	Children	of Onset	Duration		
Palpable post-auricular cervical glands	13	Day 9	2 Days		
Rash	2	Day 9	1 day		
Malaise	5	Day 9	1 day		
Arthralgia	1	Day 9	2 days		

Laboratory Studies.—Blood samples were again obtained from all the vaccinees and their siblings 46 days after vaccination. These second filter paper disc blood samples and sera were titrated, coded, in parallel with samples obtained two weeks before vaccination. The results of the haemagglutination inhibition tests showed that all the non-immune girls given

TABLE IV

Pre- vaccination	No. of Vaccinees and Siblings Grouped According to H.A.I. Test Titres 46 Days after Vaccination								
Test Titres	10	20	40	80	160	320	640		
10 20 40 80 160 320 640	53		4*	2* 1 1	3 5 1	4* 1 8 5 1* 1	2		

^{*} Vaccinated children.

RA27/3 responded to vaccination, whereas all their nonimmune unvaccinated siblings remained seronegative. The immune child given the vaccine showed no booster response and the immune siblings showed no significant rise in titre (Table IV).

Discussion

An acceptable rubella virus vaccine should cause little if any clinical reaction and should induce lasting immunity without infecting close contacts. If a vaccine were prepared from an insufficiently attenuated virus, it would give rise to an infection which was transmissible to rubella-susceptible contacts. If overattenuated the vaccine would be inadequate as an immunizing agent because of the low seroconversion rates and low postvaccination antibody titres. Although it is desirable that the rubella virus should not be excreted from the nasopharynx after vaccination, so as to preclude the chance of infecting a pregnant woman, this attribute has been observed only after vaccination with an overattenuated strain (Buynak et al., 1968). Nasopharyngeal excretion of virus therefore will occur in a proportion of subjects vaccinated with an effective vaccine strain. This characteristic is acceptable provided infection does not spread to contacts.

By vaccinating the youngest rubella-susceptible female children in large families and subjecting several rubellasusceptible siblings to intimate contact with the vaccinees, the conditions in favour of transmission of infection from vaccinees were maximal. Nevertheless, serological studies showed that infection was not transmitted by the vaccinees in any of the 19 families involving 53 rubella-susceptible contacts. The vaccine induced an immunizing infection in all the rubellasusceptible vaccinated children, with satisfactory postvaccination haemagglutinating antibody titres, though in general somewhat lower than those obtained in subjects who previously had rubella. The reactions caused by the vaccine were minimal apart from a mild degree of arthralgia for two days in an 8-year-old girl.

We wish to thank Dr. M. Flynn, County Medical Officer of Health, Westmeath, and Nurses M. O'Brien and C. Pollard for their invaluable help in the conduct of this trial.

REFERENCES

nak, E. B., Hilleman, M. R., Weibel, R. E., and Stokes, J., jun. (1968). Journal of the American Medical Association, 204, 195. Draper, C. C., and Kelly, A. (1969). British Medical Journal, 1, 177. Dudgeon, J. A., Marshall, W. C., Peckham, C. S., and Hawkins, G. T. 1969. British Medical Journal, 1, 271.

Huygelen, C., and Peetermans, J. (1967). Archiv für die gesamte Virusforschung, 21, 357.

Martin du Pan, R., Huygelen, C., Peetermans, J., and Prinzie, A. (1968). American Journal of Diseases of Children, 115, 658.

Meyer, H. M., Parkman, P. D., and Panos, T. C. (1966). New England Journal of Medicine, 275, 575.

Plotkin, S. A., Farquhar, J., Katz, M., and Ingalls, T. H. (1967). American Journal of Epidemiology, 86, 468. Plotkin, S. A., Ingalls, T. H., Farquhar, J. D., and Katz, M. (1968). Lancet, 2, 934.

Stewart, G. L., Parkman, P. D., Hopps, H. E., Douglas, R. D., Hamilton, J. P., and Meyer, H. M. (1967). New England Journal of Medicine, 276, 554.